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(54) Title: BIOSTABLE POLYURETHANE PRODUCTS

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(57) Abstract

A biostable porous polyether or polycarbonate polyurethane implant is manufactured from diphenyl methane diisocyanate, difunctional polytetramethylene ether glycol or a polycarbonate polyol, a trimerisation catalyst, a chain extender, water, a cross—linking agent, a blowing and/or gelling catalyst and a surfactant. The porous biomaterial has isocyanurate linkages and avoid content in excess of 85%. The implant may be used as an occluder or a tissue bridge.

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"BIOSTABLE POLYURETHANE PRODUCTS"

This invention relates to biostable biocompatible polyurethane implants suitable for short and long term implantation in vivo.

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Introduction

Known medical grade polyether polyurethane elastomers are susceptible to phagocyte mediated Environmental Stress Cracking (ESC). Because of this known surface phenomenon such polyether polyurethanes are not suitable candidates to provide a foam implant. Conventional polyurethane foam structures have ether linkages which are subject to oxidation induced by an inflammatory response in vivo. This is the first stage in ESC which for foam materials ultimately represents a complete loss of material and device integrity.

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There is therefore a need for a polyurethane based foam implant material which will overcome these problems.

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In particular it is an object of the current invention to provide a new class of low density biostable polymers suitable for implantation.

Statements of Invention

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According to the invention there is provided a biostable porous polyurethane foam implant material, the material containing isocyanurate linkages derived from diphenyl methane diisocyanate (MDI) and having a cross-linked structure, and a void content of at least 85%, preferably in excess of 90%, most preferably in excess of 95%, and ideally in excess of 97%.

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In particular the invention provides a biostable porous polyether or polycarbonate polyurethane article characterised in that the polyurethane is manufactured from diphenyl methane diisocyanate (MDI) with a 2, 4 MDI isomer content of less than 3%, difunctional polytetramethylene ether glycol (PTMEG) or a polycarbonate polyol, a chain extender, water, a cross-linking agent, and a trimerisation agent, a blowing and/or gelling catalyst and a surfactant. The porous biomaterial is characterised in that it has a void content in excess of 85%, is cross-linked, the hard phase contains isocyanurate linkages and the materials are biostable.

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We have surprisingly been able to produce low density polyether and polycarbonate polyurethane porous foam that is highly biostable. The biostability of the materials of the invention is especially surprising in view of the known problems with conventional polyether polyurethanes and the high surface area to volume ratio of the materials. The excellent biostability of the soft phase of the material is attributable, in part, to the method of manufacture. The material is prepared in a manner whereby the molecules in the material are virtually stress free. The stress free configuration of the molecules of the system provides the highest activation energy barrier to degradation. The materials are processed using a reaction blow moulding free rise method. The stability of the hard phase is ensured by the incorporation of highly stable isocyanurate linkages. A high level of cross-linking in the hard phase enhances its stability.

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The cross-linking agent is employed at high concentrations so as to stabilise the foam. These cross linking agents are highly reactive due to the presence of an amine linkage in their structure. They react with isocyanate at an early stage in the curing process. This helps to capture the blow and prevent blow-off.

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The trimerisation catalyst causes three isocyanates to trimerise into an isocyanurate structure. This adds additional functionality and is particularly

important because the reaction is temperature dependent. This reaction is most aggressive in the region of maximum exotherm. This corresponds to the time in the reaction when the blow is at a maximum. This additionally helps to stabilise the foam at the most critical point in the process and together with the cross-linking agent leads to rapid structure building at this time.

The high isocyanate index ensures that the isocyanate is in excess and this facilitates the trimerisation reaction and the triethanolamine reaction. The excess isocyanate provides the final cross-linking during the post cure phase of the process. At this stage in the reaction allophanate and/or biuret linkages are formed.

In a preferred embodiment the cross-linking agent is an alkanol amine. More preferably the cross-linking agent is triethanolamine.

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In a preferred embodiment the triethanolamine content is between 1% and 5% of the formulation mass. More preferably the triethanolamine content is between 1% and 3% of the mass of the formulation.

- In a preferred embodiment a trimerisation catalyst is employed. The trimerisation catalyst may be a tertiary amine, an organophosphorous compound, a metal alkyl or a carboxylate. Carboxylates are preferred and potassium acetate is the most preferred trimerisation catalyst.
- In a preferred embodiment the amount of potassium acetate is between 0.02% and 0.12% of the mass of the formulation. More preferably the potassium acetate content is between 0.06% and 0.07% of the mass of the formulation.

The invention also provides a process for preparing an implant of the invention, the processing comprising;

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preparing an isocyanate terminated prepolymer in an excess of MDI with a 4, 4 MDI isomer content of greater than 97%;

preparing a diol reaction mixture comprising a diol, a cross linking agent, a trimerising catalyst, water, a blowing and/or gelling catalyst, and a surfactant;

mixing the prepolymer and the diol;

dispensing the mixed reaction ingredients into a preheated mould which permits free rise of the foam;

post curing the foam;

demoulding the foam; and

machining the foam to form an implant.

Various other features of the invention are given in the claims.

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Brief Description of the Drawings

Fig. 1.1 is a SEM image of Polyether polyurethane biomaterial placed in a chamber with a 0.45µm filter, implanted subcutaneously in the rat animal model and explanted after 8 weeks. The integrity of the voids of the biomaterial will be noted. The structure of the voids is similar to that for the control sample (Fig. 1.2) demonstrating no evidence of ESC;

Fig. 1.2 is a SEM image of Polyether polyurethane biomaterial control sample which was not implanted in the rat model;

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Fig. 1.3 is a SEM image of Polyether polyurethane biomaterial placed in a chamber with a 0.45µm filter, implanted subcutaneously in the rat animal model and explanted after 8 weeks. As in Fig. 1.1, the integrity of the voids of the biomaterial will be noted. The structure of the voids is similar to that for the control sample (Fig. 1.2) demonstrating no evidence of ESC;

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Fig. 1.4 is a SEM image (at high magnification) of Polyether polyurethane biomaterial placed in a chamber with a $0.45\mu m$ filter, implanted subcutaneously in the rat animal model and explanted after 8 weeks. The integrity of the void of the biomaterial will be apparent. The structure of the voids is similar to that for the control sample (Fig. 1.2) demonstrating no evidence of ESC;

Fig. 1.5 is a SEM image of Polyether polyurethane biomaterial placed in a chamber with a $3.0\mu m$ filter, implanted subcutaneously in the rat animal model and explanted after 26 weeks. The integrity of the voids of the biomaterial and the cellular deposition on the biomaterial will be noted. There is no evidence of ESC;

Fig. 2.1 is a photomicrograph of the scaffold of Example 1 stained with Haemoxylin and Eosin (H&E), 12 weeks following implantation. The presence of numerous macrophage cells throughout the scaffold and the presence of blood capillaries (indicated by the arrow) in the centre of the scaffold will be noted;

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Detailed Description

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The invention provides a biostable porous polyether or polycarbonate polyurethane article characterised in that the polyurethane is manufactured from diphenyl methane dissocyanate (MDI) with a 2, 4 MDI isomer content of less than 3%, difunctional polytetramethylene ether glycol (PTMEG) or a polycarbonate polyol, a chain extender, water a cross-linking agent and a trimerisation agent. The porous biomaterials are characterised in that it has a void content in excess of 80%, is cross-linked, the hard phase contains isocyanurate linkages and the materials are biostable.

The biomaterials of this invention have unique physical and chemical characteristics that make them very suitable for in vivo implantation. The materials are especially suitable to transcatheter implantation due to the extremely high void content. Thus, the implants may be used, for example, as an occuluder or a tissue bridge.

MDI is the preferred isocyanate of this invention. The condensation of aniline with formaldehyde may produce MDI containing 2,4 and 4'4 isomers. MDI is offered in a wide variety of products, which include the 4,4 isomer 'pure' MDI product, and a variety of polymeric MDI for various different applications

Crude (polymeric) MDI is the product formed from the initial production stages where no purification has been carried out to separate various isomers and higher polymeric forms. As an example Emerck crude MDI contains the following components: 55% 4,4 and 2,4 diphenylmethane diisocyanate, 25% triisocyanate, 20% polymeric isocyanate. Due to the myriad of structures within the crude material it is preferable to use a pure grade of MDI for materials of this invention.

Pure MDI is classified as an isomer mixture of 4,4' diphenylmethane diisocyanate and 2,4' diphenylmethane diisocyanate. For the materials of this invention the 4,4' isomer is preferred. The pure MDI can be obtained as a solid, a solid in flake form or as a liquid. Pure 4,4'-MDI is sensitive to heat and will dimerise. For pure solid MDI storage below 5°C is recommended and although this will slow down the rate of dimerisation, there is still a limited time for which the pure MDI can be stored. Below 5°C a shelf life of approximately 6 months is recommended.

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For the biomaterials of this invention it is preferred that pure Flake MDI is used with an NCO content between 33.4% to 33.7%, a 2'4 MDI content <3%, a dimer content of <1%, a purity of 99.5% and a hydrolysable chlorine content of <50ppm. In an embodiment of this invention the isocyanate in the reaction chamber is in excess. More preferably the isocyanate index is between 1.03 and 1.20. Even more preferably the isocyanate index is between 1.06 and 1.16. Most preferably the isocyanate index is 1.13.

The most important characteristics of a polyol for the production of a low density foam are:

- The hydroxyl group (or amine) should be reactive to isocyanate.
 - The viscosity should be as low as possible relative to its molecular weight and functionality.
 - Preferably the polyol should have a functionality in the region of 3.

The reactivity of the hydroxyl group is dependent on whither it is a primary, tertiary, or quaternary -OH group. For example the reactivity of secondary hydroxyl end groups is several times less reactive with isocyanate than primary hydroxyl groups.

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The viscosity of the polyol needs to be as low as possible. The table below shows the viscosities of a number of commercially available polyols. The viscosity clearly depends on molecular weight and functionality. The importance of viscosity lies in the fact that the resistance to blowing is determined by the viscosity. Polyols with reduced viscosity are preferred for low density applications.

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Material	Supplier	Molecular Weight	Viscosity
PPG Diol	Lyondell	1000	110 to 220cps @
			25'C
PPG Diol	Lyondell	2245	350 to 700 cps @
			25'C
PPG Triol	Lyondell	1070	400cps @ 25'C
PTMEG Diol	Du Pont	1000	250 to 320cps
			@40'C
PTMEG	Du Pont	2000	950-1450 cps @
			40'C
CX5510	Nissei	1000	1610cps @50'C
(Polycarbonate	Chemicals		
diol)			
CX5520	Nissei	2000	10,400 cps @50'C
(Polycarbonate	Chemicals		
Diol)			

The functionality of the polyol is crucial to stabilising the expanded formulation. High functionality increases the rate at which molecular weight is developed. This molecular weight increase prevents the internal pressure of the gas from blowing the foam apart. This molecular weight increase also prevents the foam from shrinking as the blowing gas is liberated during the cell opening phase.

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The invention provides a range of biostable materials based on these polyols. In particular low density foams using 'tertiary carbon' free polyols are provided.

Multifunctional polyols are usually prepared using a trifunctional initiator, such as glycerol or trimethylolpropane. These materials contain tertiary carbons and would be susceptible to attack and degradation by phagocytic agents. The absence of these linkages enhances the biostability of the materials. Difunctional polyols are thus advantageous from an implantation biostability stand point. This is especially important where the implant is of high surface area and low density.

The lack of functionality and the high relative viscosity of polytetramethylene ether diols and polycarbonate diols make the blowing and stabilisation of low density foams from these materials extremely difficult. The complexity is added to by the need of the resulting foams to have excellent biostability characteristics.

High density PTMEG foams have been produced for applications in shoe manufacture. Du Pont have produced PTMEG microfoams with a density range from 250kg/m³ to 300kg/m³. Low density foam is defined per this invention as having a void content in excess of 80%. Soft microcellular polycarbonate urethane foams are also described by Pinchuck EP461 375.

The invention provides a range of low density PTMEG and polycarbonate polyol based polyurethanes with properties suitable for long term biomedical implantation.

The PTMEG materials of this invention capture the excellent resilience properties of PTMEG and overcome the biostability issues of conventional polyether urethanes.

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PTMEG is the preferred polyether polyol of this invention. PTMEG is manufactured from Polytetrahydrofuran (PTHF) by a ring opening polymerisation technique. The linearity of PTMEG increases the difficulty in preparing low density 3 dimensional porous matrices with this material. The linearity results from the fact that no polyfunctional initiator is used during the polymerisation step. This is unlike the polymerisation of other polyethers used in polyurethane foam manufacture that can use a trifunctional or higher functionality initiator to increase the functionality of the polyether polyol.

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The PTMEG polyols used in this invention have molecular weights of 400 – 5000, more preferably between 500 –2000.

PTMEG is a white waxy solid material at low temperatures and has a melt temperature ranging from 11°C to 43°C depending on the molecular weight of the PTMEG.

The quality of PTMEG starting materials is important for the biomaterials of this invention. The preferred molecular weights of the polyols for this application are 650, 1000 and 2000 with minimum polydispersity. Ideally the PTMEG starting materials will have characteristics as outlined below;

Characteristic	650 molecular	1000 molecular	2000 molecular
	weight PTMEG	weight PTMEG	weight PTMEG
Molecular weight	600 - 700	950 – 1050	1900 – 2100
OH value	160 - 187	112 - 118	53 –59
Water content	<0.015 % wt	<0.015 % wt	<0.015 % wt
Viscosity @40°C cP	100 - 200	260 - 320	950 – 1450
Antioxidants	<0.1%	<0.1%	<0.1%

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The development of molecular weight during the foaming step is a critical aspect of the invention. The importance of this area results from the following limitations of PTMEG as a foaming material:

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- PTMEG has lower reactivity than standard foaming polyols.
- PTMEG is commercially available with a functionality of 2. Standard PPG foaming polyols have functionalities above 3 and as high as 8.
- PTMEG is significantly more viscous than standard PPG based foaming polyols

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The preparation of PTMEG and polycarbonate foams with void contents in excess of 90%, 95% and 97% is also unexpected. This has been achieved to spite the limitations of difunctional, high viscosity polyols as a foaming ingredient. A strong blowing catalyst is used to deal with the high relative viscosity of the polyol. This pushes the blowing reaction in the early stages of foaming, while the molecular weight is low. Normally this approach would exacerbate the lack of functionality of the polyol and the foam would be blown apart towards the end of the blowing phase. However we have introduced mechanisms for developing the molecular weight to deal with the fast blowing reaction. These mechanisms include the use of a reactive cross-linking agent and the incorporation of a trimerisation catalyst. The cross-linking agent provides functionality early in the blowing process. The inclusion of a trimerisation catalyst ensures cyclic isocyanurates groups form during the later part of the blowing phase. The incorporation of isocyanurates introduces a further element of cross-linking and provides additional stability to the foam. The trimerisation catalyst lowers the temperature for the formation of isocyanurates. The rate of isocyanurate formation increases with temperature and is thus linked to the exotherm, the material and the mould temperatures.

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The polycarbonate diols of the invention have the following general characteristics:

- The polycarbonate diols are difunctional polyols.
- They are free of tertiary carbons.
- They have molecular weights of from 400 to 5000.
- They are linear unbranched polycarbonate diols.

The diols of the invention are generally manufactured by a reaction of a carbonate with a diol or a mixture of diols. Examples of the polycarbonate polyols used in the invention are disclosed by Greco EP 0 533275, Lia U.S. 4,131,731 and Funakoshi EP 0 754 714 which are included herein in their entirety for reference. Preferred diols are linear aliphatic diols. Preferably the polycarbonate has between 4 and 20 carbons between carbonate linkages. More preferably between 4 and 10.

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The materials and processes of the invention allow for the early expansion of the foam in a low viscosity state. As the formulation reaches full blow molecular weight is developed quickly and this stabilises the expanded material. The rapid development of molecular weight at the time of full blow is achieved by building three separate types of functionality into the formulation.

- 1. The incorporation of a reactive cross-linking agent into the formulation provides tri-functionality early in the process.
- 2. A trimerisation catalyst generates trifunctional isocyanurates at the time of maximum blow. This occurs because of the temperature dependent nature of the trimerisation reaction and it is accelerated by the exotherm generated in the blowing reaction. The materials and mould temperatures are important in optimising this portion of the reaction.
- 3. The excess isocyanate reacts with urethane linkages already formed. This provides cross-links in the later stages of the process.

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It is the combination of these three events that makes the stabilisation of the foams of the invention possible.

The cross-linking agent is a reactive component that has a functionality of three. 5 It forms covalent bonds when reacted with the diisocyanate, resulting in the formation of a three dimensional network within the material.

Cross-linking agents can be grouped into three classes, alcohols, amines and alkanol amines.

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The alkahols include glycerol, trimethylol propane, and sorbitol. As all of these contain tertiary carbons they are not preferred for the manufacture of the biostable foams of the invention.

The amines include MBOCA (3,3'-dichloro-4,4'-diamino-diphenylmethane) and 15 diethylenetriamine.

Alkanol amines include ethanol amine, diethanolamine and triethanolamine.

Alkanol amines are preferred per this invention. A characteristic of these cross-20 linking agents is that they function as catalysts, thus they maybe classified as reactive catalysts. A high purity grade is preferred (99+%).

Water is employed as the blowing agent in the materials of the invention. The quality of the water used in this formulation is important and is controlled. HPLC grade water with a residue on evaporation of < 0.0005% may be used. However, more preferably deionised water with no detectable oxidisable substances, chlorides, nitrates sulphates and ammonium, < 0.001% of residue on evaporation, pH 5 - 8 and endotoxin levels < 0.5EU/ml is used. High pyrogen levels in starting materials of products destined for human implantation should

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be avoided. The water content of the polyol resin component ranges from 2-5 php (parts per hundred polyol) depending upon the void content of biomaterial required.

- 5 There are two main classes of catalysts in the manufacture of polyether polyurethanes.
 - Organic tin catalysts primarily accelerate the gelling reaction
 - ◆ Tertiary amine compounds to promote both the gelling and the blowing reactions

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Amine catalysts are preferred for this invention. Depending on the structure of the amine either the gelling or blowing reaction is favoured. By careful selection of the catalysts a desirable rate between the gelling and blowing reactions can be achieved to yield a low density material.

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Amine catalysts including but not limited to triethylenediamine, N,N,N'N'-tetramethylhexamethylenediamine, bis(2-dimethylaminoethyl)methyl amine, N-(2-dimethylaminoethyl)-N'-methylpiperazineand N-(2-dimethylaminoethyl)ether can be used.

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The preferred catalyst system for this invention is Desmorapid PP (From Whitchem), which is a combination of bis(2-dimethylaminoethyl)methyl amine and N-(2-dimethylaminoethyl)-N'-methylpiperazine, with triethylenediamine. The resulting combination yields a strong blowing reaction.

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A further class of catalysts is based on carboxylates. These promote the trimerisation reaction of the isocyanate and are critical to the formation of the stable foams of the invention. The inclusion of a trimerisation catalyst ensures cyclic isocyanurates groups form during the later part of the blowing phase. The incorporation of isocyanurates introduces a further element of cross-linking and

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provides additional stability to the foam. The trimerisation catalyst lowers the temperature for the formation of isocyanurates. The rate of isocyanurate formation increases with temperature and is thus linked to the exotherm, the material and the mould temperatures.

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The trimerisation catalyst causes three isocyanates to trimerise into an isocyanurate structure. This adds additional functionality and is particularly important because the reaction is temperature dependent. This reaction rate is fastest in the region of maximum exotherm. This corresponds to the time in the reaction when the blow is at a maximum. This additionally helps to stabilise the foam at the most critical point in the process and together with the cross-linking agent leads to rapid structure building at this time.

The high isocyanate index ensures that the isocyanate is in excess and this provides raw material for the trimerisation reaction and the cross linking reaction.

In a preferred embodiment a trimerisation catalyst is employed. The trimerisation catalyst may be a tertiary amine, an organophosphorous compound, a metal alkyl or a carboxylate. Carboxylates are preferred and potassium acetate is the most preferred trimerisation catalyst.

In a preferred embodiment the amount of potassium acetate is between 0.02% and 0.12% of the mass of the formulation. More preferably the potassium acetate content is between 0.06% and 0.07% of the mass of the formulation.

Catalysts such as potassium acetate, sodium acetate and the like are added to the resin in a solvent carrier.

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The function of the surfactant is to control the formation and growth of the gas bubbles and with it many processing and final properties of the scaffold.

In the emulsification of the raw materials, surfactants allow thermodynamically incompatible components of a polyurethane foam formulation to mix. The concentration of gas bubbles formed during the foaming is strongly dependent on the characteristics of the surfactant used.

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During the rise of the foam the main demand on the surfactant is the stabilisation of the expanding liquid mixture. The surfactant reduces the surface tension of the liquid mixture and therefore reduces the energy the system needs to facilitate cell formation and growth.

During the rise the viscosity increases gradually within the material. The structural stability increases with this viscosity increase and the structure becomes more self-supporting. As the blow reaches its maximum the spherical voids start to impinge on one another. The surfaces of infringement become locally flattened. The surfactant plays a critical role at this stage. By controlling the surface tension some of the flattened surfaces of the impinging voids rupture. As these rupture the material in the voids reflows into the main arch.

A typical silicone surfactant generally has a siloxane backbone formed by dimethylsiloxane units. Polyether groups and/or additional modifications can be attached to the siloxane backbone. Suitable surfactants include BF 2270, BF 8002 from Goldschmidt A.G. Surfactant quantities from 0.30% to about 2% weight percent based on polyol resin may be suitable. More preferably the weight % of the surfactant is between 0.5% and 1.2%.

Chain extenders are low molecular weight difunctional compounds. During the reaction hard segments are formed by the reaction of the diisocyanate with the

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low molecular weight chain extender. Water used as a blowing agent can also be considered a chain extender as it is a low molecular weight difunctional component. Diols are the preferred chain extender.

5 1-4-butane diol is a preferred chain extender. A high purity grade is required (99+%), as impurities such as 2-methyl butane-1-4-diol should be avoided.

Other diol chain extenders that can be considered are, but not limited to, ethylene glycol, propylene glycol, 1-6-hexane diol, diethylene glycol, dipropylene glycol.

The materials of the invention are manufactured by a quasi-prepolymer method. In the production of an isocyanate quasi prepolymer the diisocyanate is reacted with a portion of polyol to form a relatively low viscosity prepolymer which is then subsequently reacted with the remainder of the polyol, water, various catalysts, surfactants and chain extenders and other ingredients.

The key outputs from the preparation of the prepolymer process are the NCO content of the prepolymer and the resulting viscosity of the prepolymer. These outputs are controlled in the process by controlling the temperature and the method of preparation. The prepolymer is preferably prepared at a temperature between 70°C and 80°C. This provides the best control of viscosity and minimises side reactions. Possible undesirable side reactions at this stage include the formation of allophanates and trimerisation.

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The method in which the prepolymer is prepared has an affect on the viscosity of the prepolymer. If the liquid polyol is added to isocyanate a lower viscosity prepolymer results than if the isocyanate is added to the liquid polyol. This can be explained in terms of the molecular weight distribution within the prepolymer. The addition of the liquid polyol to the isocyanate is the preferred

method. The lower the viscosity of the prepolymer the lower the resulting density of the formulation.

The polyol is prepared by adding the required quantity of polyol (of the molecular weight required) to a round bottomed flask, placing in the heating mantle and melted. The appropriate volume of water is added and mixed. Appropriate quantities of the remaining components (chain extender, blowing agent, catalysts, surfactant and cross linking agents) are added to the reaction flask and mixed thoroughly for 25-30 minutes using a moderate shear rate.

The amount of material required too manufacture even the largest implant or scaffold is only tens of grams. The smallest scaffolds per this invention are only a fraction of a gram. In a preferred embodiment the reaction ingredients are metered using precision gear pumps which can deliver an accuracy of 0.01g.

The materials of the invention are metered into a mixing chamber.

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The polyol mixture and isocyanate resin are aggressively mixed in a mixing chamber. The aggressive mixing causes the two incompatible phases to be interspersed. Mixing is crucial to the processes of the invention as it brings the reactive sites to come into close proximity and this facilitates the reactions.

Shot mass per this invention varies from as little as 1 gram. More typically the shot size varies from 2g to 10g. More typically from 3-5g.

Mixing is an important variable in controlling the size and distribution of the voids. The greater the level of material mixing the smaller the diameter of the voids.

In the design of a mixing chamber it is desirable to generate shear and turbulence. This ensures that there is homogeneity locally in the chamber and at different points in the chamber. Shear mixing is measured indirectly by the

speed of the mixer or the shear rate or the relative velocities of the components of the mixing unit or the velocity gradient. The degree of turbulence in the chamber is dependent on the design of the mixing chamber and is very difficult to quantify.

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The mixing process is characterised in that:

- The mixing chamber is circular in cross-section
- An annular space is provided between the chamber outer wall and an inner relative rotational element.
- The rotational element rotates at speeds in excess of 1000rpm, preferably between 2000rpm and 6000rpm.
 - The rotational element has slot features, and/or fin features and/or hole features and/or raised features so as to generate shear stresses and turbulence in the mixing chamber.

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The dispensing step is characterised in that the material is ejected out of the mixing chamber and into a mould. The conditions of the mould facilitate a phenomenon known as "free rise". Free rise describes a process whereby the mixed products are permitted to expand without limitation. The mixed reactants are dispensed into a mould that facilitates at least a nine fold volumetric expansion and as much as a 50 fold expansion of the biomaterial. The mould is vented to allow for free rise and is placed preferably in a carbon dioxide rich oven, which assists in the stabilisation of the materials physical structure. The temperature of the mould should be in excess of the temperature of the reactants. Preferably the mould temperature is between 80°C and 90°C. The mould should be manufactured from a material, which is non-reactive to isocyanate. PTFE, silicone polypropylene and POM are exemplary mould materials.

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The low density polyurethane biomaterial is placed in a carbon dioxide oven after dispensing into the mould for a minimum of 1 hour. The oven temperature is between 80°C - 90°C but preferably at 80°C. This process is important in that it helps prevent shrinkage of the material and allows virtually all reactive sites within the structure to react.

Reticulation of the scaffold can be carried out by crushing. This optional step increases the number of pores per void. It has the disadvantage that reticulated pores will tend to be more irregular in shape.

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It should be obvious to one skilled in the art that decreasing the density of the material causes an increase in the surface area. When the material is in the shape of a low density foam, or cellular material the surface area becomes especially large.

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The materials of this invention are two phase materials. The hard phase is generated by the reaction of MDI with small chain molecules and through trimerisation. Typically, the small chain molecules are diols, diamines, alkanol amines or water. Preferably the chain extenders are diols or water. The second phase of the material is referred to as the soft phase and is composed PTMEG or a polycarbonate polyol. The two phases differ in cohesive energy density.

The soft phase typically has a cohesive energy density in the region of 20-22 MPa^{1/2} while the hard phase typically has a cohesive energy density in the region of 24 -28MPa^{1/2}. This allows the two phases separate into distinct domains. This phase separated domain structure confers optimal biocompatibility characteristics to the material.

Phase separation is enhanced by achieving most of the crosslinking in the hard phase. The use of a difunctional polyol is preferred.

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Preferably the hard segment content is from 35 % to 65%. More preferably the hard segment content is from 35% to 55%. More ideally the hard segment content is from 40 to 50%. The Hard Segment Content is defined as follows:

5 {M.(isocyanate) + M.(chain extender) + M.(crosslinker) + M.(water) - M.(CO2) } / {M.(isocyanate) + M.(chain extender) + M.(crosslinker) + M.(water) + M.(polyol) - M.(CO2) } Where M is the mass of a material used in or generated by the reaction.

The incorporation of isocyanurate linkages in the hard phase contributes strongly to stability of the hard phase. These cyclic structures are highly stable. The incorporation of isocyanurates into a soft tissue contacting material is a unique feature of the invention.

The materials of this invention can be manufactured to indentation hardness which match the compliance of tissues in given applications. The materials of example 1 and 2 have characteristics which approximately match those of tissue. An important advantage of these highly porous foams in tissue applications is that the indentation force does not increase rapidly with the level of indentation. Harder grades of material are easily prepared using already established principles.

Examples

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The unique physical and biostability features of the polyether polyurethane urea isocyanurate and polycarbonate polyurethane urea isocyanurate biomaterials of this invention will be demonstrated by the following examples.

Example 1

For preparation of porous biostable polytetramethylene oxide polyurethane urea isocyanurate biomaterial, a polyol resin and an isocyanate pre-polymer were prepared. In the preparation of the polyol resin the following raw materials were

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added to a heated round bottom flask and mixed at 50 - 60°C for a minimum of 25 - 30 minutes.

Raw material	Quantity (php)	
Terathane (MW 1000) ¹	100	
Triethanolamine ²	4.6	
Water ²	2.56	
1,4 Butanediol ²	8.05	
BF 2270 ³	1.0	
RC Catalyst 105 ⁴	2.96	
Desmorapid PP ⁵	0.34	
Kac/Deg ⁶	0.73	

¹PTMEG (Du Pont), ²Sigma Aldrich, ³Th GoldSchmidt, ⁴Triethylenediamine (Rhein Chemie), ⁵Whitchem, ⁶Potassium acetate (Sigma Aldrich)

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An isocyanate pre-polymer with an NCO content of 15.6% was prepared by charging flake MDI (Desmodur 44M flakes from Bayer, MDI-2,4' isomer content of 1.37%) into a heated round bottom flask and allowing the flakes to melt. Upon complete melting of the MDI, the PTMEG (Terathane MW 1000) was added and the reaction mixture was stirred at 70 - 80°C for 60 to 90 minutes. Dry nitrogen was purged through the reaction flask at all times to eliminate moisture from the reaction vessel.

15 The polyether polyurethane biomaterial was prepared using metering, mixing and dispensing equipment manufactured by 2KM (Germany). The processing temperature of the materials was 40°C and mixing was achieved at a speed of 5000 RPM. The materials were dispensed into a pre heated silicone mould of temperature 80 – 90°C at an isocyanate index of 1.13. The combined shot size of the materials was 2.2g. The mould was placed in a CO₂ vented oven at a

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temperature of 90°C for a minimum time period of 90 minutes. The biomaterial was reticulated using a hand press.

Samples were taken post curing to perform the tests listed below.

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Results.

The calculated hard segment content of this material was 42.6 %

A sample of the foam was cut using a cutting blade into a rectangular block, ensuring no skin was present on the material. The dimensions were recorded using a calliper and the weight was recorded on an electronic balance.

The void content was calculated as 90% based on a solids density of 1200 Kg/m³.

- The indentation hardness of the above material was evaluated per ISO standard 2439 with the following deviations
 - indenter diameter of 10mm.
 - sample thickness of 10mm,
 - sample diameter of 20mm.
- The sample was prepared by first cutting the sample with a cutting blade and then lasering the appropriate cylinder from the piece of material.

The indentation hardness recorded were 1.14N at 25%, 1.3N at 40% and 4.14N for 65%.

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Biostability

Samples of this biomaterial were placed in sealed acrylic chambers and implanted in the subcutaneous cavities of rats. The acrylic chambers were sealed with $0.45\mu m$ and $3.0\mu m$ filters to allow fluids have contact with the biomaterial.

The samples were explanted at a number of time points up to 6 months post

implantation. The biomaterial samples were examined by Scanning Electron Microscopy (SEM). The images provided in Figures 1.1, 1.3, 1.4 and 1.5 demonstrate that the structural integrity of the biomaterial is comparable to the biomaterial sample that was not subjected to the in vivo environment (Figure 1.2). Histological analysis of this biomaterial (Figure 2.1) demonstrates the presence of a massive and persistent macrophage response. However, despite the presence of a large number of macrophage no evidence of degradation was observed in any of the explanted samples, as demonstrated.

- This biomaterial was also implanted in the gluteal muscle of rats and left for up to 6 months. The histological analysis conducted on the explants indicated that;
 - ♦ The biomaterial was covered with tissue within 4 weeks of implantation.
 - Angiogenesis was clearly observed within the bulk of the biomaterial 4 weeks following implantation. This anchors the implant in relation to the surrounding tissue.
 - ♦ No evidence of biomaterial degradation was observed.

The biomaterial scaffold was implanted in the vasculature of a rabbit for a period of up to 3 months. Again no degradation of the material was observed.

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Example 2

For preparation of porous biostable polytetramethylene oxide polyurethane urea isocyanurate biomaterial, a polyol resin and an isocyanate pre-polymer were prepared. The polyol is prepared per example 1 using the following chemicals

Raw material	Quantity (php)	_
Terathane (MW 1000) ¹	100	
Triethanolamine ²	4.6	
Water ²	3.0	
1,4 Butanediol ²	8.0	
BF 2270 ³	1.2	
RC Catalyst 105 ⁴	0.52	
DABCO BL11 ⁵	0.13	
Kac/Deg ⁶	0.73	

¹PTMEG (Du Pont), ²Sigma Aldrich, ³Th GoldSchmidt, ⁴Triethylenediamine solution (Rhein Chemie), ⁵Air Products, ⁶ Potassium acetate solution (Sigma Aldrich)

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An isocyanate pre-polymer with a NCO content of 25.0% was prepared as per example 1 using PTMEG polyol (Terathane MW 650).

The polytetramethylene oxide polyurethane urea isocyanurate biomaterial was prepared per the method of example 1 at an isocyanate index of 1.13 with the exception that the combined shot size of the components was 1.3g

Results.

The calculated hard segment content of this material was 51.9 %

A rectangular block was cut as per example 1 and weighed. The void content was calculated as 95% again based on a base line density of 1200Kg/m³.

Example 3

For preparation of a porous biostable polycarbonate polyurethane urea isocyanurate biomaterial, a polyol resin and an isocyanate pre-polymer were prepared.

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The polyol is prepared per example 1 using the following chemicals.

Raw material	Quantity (php)
Polycarbonate CX 5510 (MW 1000) ¹	100
Triethanolamine ²	3.6
Water ²	3.0
BF 2270 ³	1.2
RC Catalyst 1054	1.66
DABCO BL 11 ⁵	0.8
Kac/Deg ⁶	0.73

¹Nissei Chemical Company, Japan. ²Sigma Aldrich ³Th GoldSchmidt ⁴Triethylene diamine solution (Rhein Chemie) ⁵Air Products ⁶Potassium acetate solution (Sigma Aldrich)

The isocyanate pre-polymer was prepared as per example 1 using polycarbonate CX 5510 (MW 1000) to produce a prepolymer with an isocyanate content of 15.6%

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Polycarbonate CX5510 is a random co-polymer comprising hexamethylene carbonate and pentamethylene carbonate sequences of 1000MW.

The materials were mixed at an isocyanate index of 1.13 in an open top reaction vessel. The temperature of the materials was 40°C and a mixing speed of 2200RPM was achieved. The combined shot size of the materials was 52g.

Results.

A rectangular block was cut as per example 1 and weighed. The void content was calculated as 88% based on a solids density of 1200Kg/m³.

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Indentation hardness values were measured using the same procedure as per example 1. The following results were obtained: 2.3N at 25%, 3.2N at 40% and 7.8N for 65%.

The invention is not limited to the embodiments hereinbefore described which may be varied in detail.

Claims

- 1. A biostable porous polyurethane foam implant material, the material containing isocyanurate linkages derived from diphenyl methane diisocyanate (MDI), and having a cross-linked structure, and a void content of at least 85%.
- 2. An implant as claimed in claim 1 wherein the material has a void content in excess of 90%.
 - 3. An implant as claimed in claim 1 wherein the material has a void content in excess of 95%.
- An implant as claimed in claim 1 wherein the material has a void content in excess of 97%.
 - 5. An implant as claimed in any preceding claim wherein the material is manufactured from

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diphenyl methane diisocyanate (MDI) with a 2,4 MDI isomer content of less than 3%;

a linear, long chain diol which is free of tertiary carbon linkages;

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water;

a cross-linking agent;

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a trimerisation catalyst;

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a blowing and/or gelling catalyst; and

a surfactant.

- 5 6. An implant as claimed in claim 5 wherein the diol is polytetramethylene ether glycol (PTMEG).
 - 7. An implant as claimed in claim 5 wherein the diol is a polycarbonate diol.

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- 8. An implant as claimed in claim 7 wherein the polycarbonate diol is a reaction product of one or more diols with a carbonate monomer.
- 9. An implant as claimed in any of claims 5 to 8 wherein the diol molecular weight is between 400 and 5000.
 - 10. An implant as claimed in claim 9 wherein the diol molecular weight is between 500 and 2500.
- 20 11. An implant as claimed in any of claims 5 to 10 wherein the trimerisation catalyst is a carboxylate.
 - 12. An implant as claimed in claim 11 wherein the trimerisation catalyst is a potassium acetate.

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13. An implant as claimed in claim 12 wherein potassium acetate is present in the reaction formulation in an amount of from 0.02% to 0.12% by mass of the formulation.

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- 14. An implant as claimed in any of claims 5 to 13 wherein the cross-linking agent is present in the reaction formulation in an amount of from 1% to 5% by mass.
- 5 15. An implant as claimed in claim 14 wherein the cross-linking agent is trialkanol amine.
 - 16. An implant as claimed in claim 15 wherein the cross-linking agent is triethanolamine.

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- 17. An implant as claimed in any preceding claim wherein the isocyanate index of the reaction formulation is from 1.03 to 1.20.
- 18. An implant as claimed in claim 17 wherein the index is approximately 1.13.
 - 19. An implant as claimed in any of claims 5 to 18 wherein the reaction formulation includes a chain extender.
- 20. An implant as claimed in claim 19 wherein the chain extender is a linear aliphatic diol.
 - 21. An implant as claimed in claim 20 wherein the linear aliphatic diol is 1, 4 butane diol.

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22. An implant as claimed in any of claims 19 to 21 wherein the chain extender is present in the formulation in an amount of less than 7% by mass.

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23.	An implant as claimed in claim 22 wherein the chain extender is present
	in the formulation in an amount of less than 4% by mass.

- 24. An implant as claimed in any of claims 5 to 23 wherein water is present in the reaction formulation in an amount of from 0.6% to 1.8% by mass.
 - 25. An implant as claimed in any preceding claim wherein the implant has a hard segment content of from 35 to 65%.
- 10 26. An implant as claimed in claim 25 wherein the implant has a hard segment content of from 35 to 55%.
 - 27. An implant as claimed in claim 25 or 26 wherein the implant has a hard segment content of from 40 to 50%.

28. An implant as claimed in any preceding claim in the form of an occluder.

- 29. An implant as claimed in any of claims 1 to 27 in the form of a tissue bridge.
- 30. A process for preparing a biostable polyurethane implant as claimed in any of claims 1 to 29 comprising the steps of:-
- preparing an isocyanate terminated prepolymer in an excess of MDI with a 4, 4 MDI isomer content of greater than 97%;

preparing a diol reaction mixture comprising a diol, a cross-linking agent, a trimerising catalyst, water, a blowing and/or gelling catalyst, and a surfactant;

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mixing the prepolymer and the diol;

dispensing the mixed reaction ingredients into a preheated mould which permits free rise of the foam;

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post curing the foam;

demoulding the foam; and

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machining the foam to form an implant.

- 31. A process as claimed in claim 30 wherein the foam is post cured in a carbon dioxide rich environment.
- 15 32. A process as claimed in claim 30 or 31 including the step of preheating the mould prior to dispensing of the mixed reaction ingredients.
 - 33. A process as claimed in any of claims 30 to 32 including the step, after demoulding, of crushing the foam to increase the open cell content of the foam.
 - 34. A process as claimed in any of claims 30 to 33 wherein the prepolymer is prepared from a prepolymer reaction mixture at a temperature of from 70 to 80°C.

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- 35. A process as claimed in any of claims 30 to 34 wherein the prepolymer reaction mixture is reacted for a period of from 1 to 2 hours.
- 36. A process as claimed in any of claims 30 to 35 wherein the prepolymer reaction mixture is stirred continuously under a dry inert atmosphere.

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37. A process as claimed in any of claims 30 to 36 wherein during moulding the mould temperature is maintained at a temperature of greater than 50°C.

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38. A process as claimed in claim 37 wherein the mould temperature is from 80 to 90°C.

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- 39. A process as claimed in any of claims 30 to 38 wherein the post curing is carried out at a temperature of at least 50°C for a period of at least 30 minutes.
- 40. A process as claimed in claim 39 wherein the post-curing is carried out at a temperature of approximately 80°C.

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- 41. A process as claimed in any of claims 30 to 40 wherein the shot size is less than 10g.
- 42. A process as claimed in claim 41 wherein the shot size is less than 5g.

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43. A process as claimed in claim 41 or 42 wherein the shot size is less than 3g.

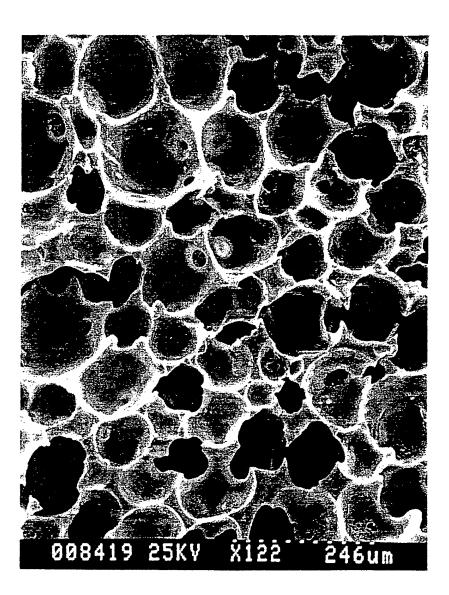


Fig 1.1

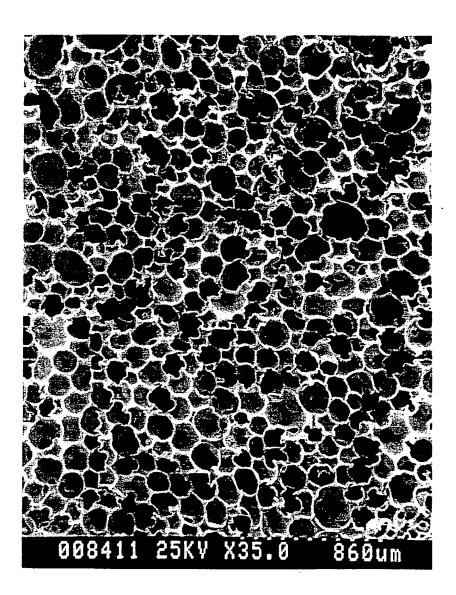


Fig 1.2

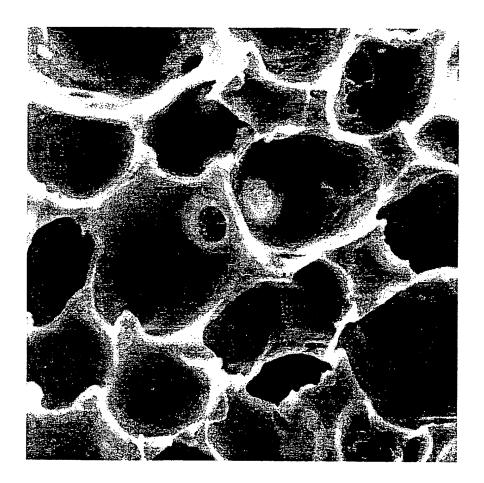


Fig 1.3

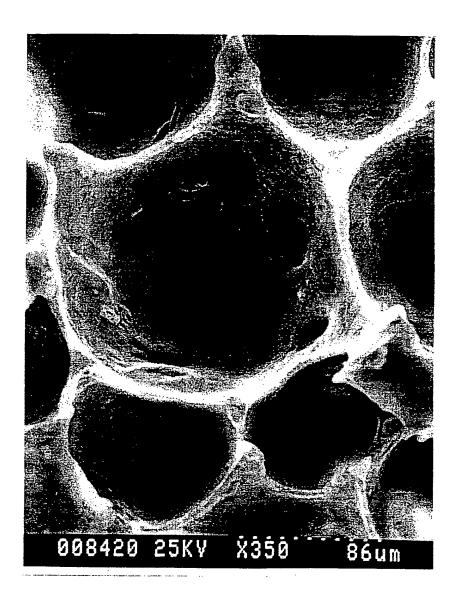


Fig 1.4



Fig 1.5

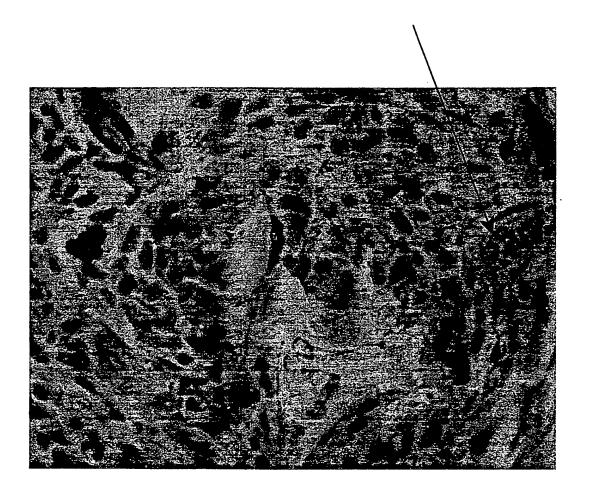


Fig. 2.1

INTERNATIONAL SEARCH REPORT

Inte. Jonal Application No PCT/IE 00/00056

A. CLASSIFICATION OF SUBJECT MATTER IPC 7 A61L27/18 A61L27/56

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols) IPC 7 A61L

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

EPO-Internal, CHEM ABS Data, MEDLINE, EMBASE, BIOSIS

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X Further documents are listed in the continuation of box C.	Patent family members are listed in annex.
Special categories of cited documents: "A" document defining the general state of the art which is not considered to be of particular relevance "E" earlier document but published on or after the international filing date	"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention "X" document of particular relevance; the claimed invention
"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another	cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone "Y" document of particular relevance; the claimed invention
citation or other special reason (as specified) "O" document referring to an oral disclosure, use, exhibition or other means "P" document published prior to the international filling date but later than the priority date claimed	cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art. *&* document member of the same patent family
Date of the actual completion of the international search	Date of mailing of the international search report
27 July 2000	03/08/2000
Name and mailing address of the ISA	Authorized officer
European Patent Office, P.B. 5818 Patentlaan 2 NL – 2280 HV Rijswijk Tel. (+31–70) 340–2040, Tx. 31 651 epo nl, Fax: (+31–70) 340–3016	Diederen, J

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